A MONTHLY, OPEN ACCESS, PEER REVIEWED (REFEREED) INTERNATIONAL JOURNAL Volume 04, Issue 08, August 2025

# Effect of salinity stress on seedling growth of *Vigna mungo* (L.) Hepper Dr. Seema Anand<sup>1</sup>

<sup>1</sup>Associate Professor, Department of Botany, S. V. College, Aligarh, U.P. (India)

Received: 20 August 2025 Accepted & Reviewed: 25 August 2025, Published: 31 August 2025

# **Abstract**

Salinity reduces urad bean (Vigna mungo) radicle and root elongation, delays and inhibits hypocotyl elongation and mobilization of reserves from the cotyledons to the embryo axis. Fresh and dry masses and water content of the embryo axes were reduced. Under salinity, a net leakage of K to the media increased with time and increasing NaCl concentrations. Sugars present in the cotyledons of seeds were of primary importance for growth of the embryo axis up to 18 h after sowing whereas breakdown of starch by amylase contributed later, the contribution being delayed and reduced with increasing NaCl concentration. Even when amylase activity in the cotyledons was progressively reduced with increasing NaCl concentration, the increasing contents of soluble sugars in the cotyledons indicated that sugars were not limiting for mung bean seedling growth under salinity. Salinity stress is a major environmental factor limiting crop productivity worldwide. This research aims to investigate the effects of salinity stress on the germination of urad bean (Vigna mungo) seeds and explore potential mitigation strategies to enhance germination rates under saline conditions. The objective of our research was to study the effect of salinity stress on urad beans (*Vigna mungo*) germination by observing the result of increasing salt concentrations on the germination of mung bean seeds. The primary aim of this research is to investigate the effect of salinity stress on the germination of urad bean (Vigna mungo) seeds. This study focuses on evaluating key germination parameters, including germination percentage, germination rate, and mean germination time, under varying levels of salt concentration.

**Keywords**: Salinity, seedlings, germination, salt concentration.

# Introduction

Germination and early seedling stages are crucial in the plant life cycle, so it is important to identify and ideal growth medium to ensure crops are efficiently yielded to meet increasing global demand. When environmental conditions cannot support healthy growth, seeds enter a dormant stage to avoid premature germination and will continue to germinate if conditions become ideal again.

Soil salinity is a significant environmental challenge that threatens global agricultural productivity, particularly in arid and semi-arid regions. It is estimated that over 20% of the world's cultivated land and nearly half of all irrigated land are affected by salinity, posing serious risks to food security. Further in the life cycle, the most typical effect of salt stress on plant is stunted growth as salinity reduces cell division and the synthesis of RNA, DNA, and proteins. Salinity stress arises from excessive accumulation of soluble salts, primarily sodium chloride (NaCl), in the soil. These salts reduce the soil's water potential, making it difficult for plants to absorb water and nutrients. Additionally, high salt concentrations lead to ion toxicity and oxidative stress, further impairing plant growth and development.

Vigna mungo is an important legume crop known for its high nutritional value, rich in protein, fibre, vitamins, and minerals. It is widely cultivated in Asia, Africa, and South America due to its adaptability to various climates and its role in sustainable agriculture as a nitrogen-fixing plant. Despite its resilience to some environmental stresses, mung bean is particularly vulnerable to saline conditions. Salinity stress adversely affects key physiological processes such as water uptake, enzyme activity, and energy metabolism during germination. As a result, poor germination rates and stunted seedling growth are common under saline environments, limiting overall crop yield.

A MONTHLY, OPEN ACCESS, PEER REVIEWED (REFEREED) INTERNATIONAL JOURNAL Volume 04, Issue 08, August 2025

Understanding the impact of salinity stress on mung bean germination is crucial for developing effective strategies to improve crop performance in affected regions.

This study investigates the effects of varying salinity levels on mung bean seed germination, examining key physiological and biochemical changes. In addition, the study explores potential mitigation strategies, such as antioxidant treatments and nutrient enrichment, to improve seed germination and seedling vigour under saline conditions. By identifying practical solutions to minimize salinity-induced damage, this research aims to support sustainable agricultural practices and improve food security in saline-prone regions. Various researchers have explored the effects of salinity on *Vigna mungo*, highlighting its influence on germination rate, seedling vigour, and overall plant health.

This review presents studies conducted by prominent scientists investigating the impact of salt stress on urad bean germination and its medicinal and economical uses, such as, Jeevan Suresh Dhumal and friends (2019) studied that the bioactive compound is secondary plant metabolites eliciting pharmacological or toxicological effects in man and animals. Legumes are a valuable source of proteins and nutrients for the majority of the world population. Vigna mungo is one of the important legume crops extensively cultivated in India and other parts of the world. Pulses and legumes have been gaining interest because they are an excellent source of bioactive compounds. Fatima Khan et al (2021) studied the medicinal properties of Vigna mungo and observed that it is rich in flavonoids, iso -flavonoids, phytoestrogens, phenolic acids, enzymes, fibres, starches, trypsin inhibitors, phytic acid, lectins, saponins, tocopherols, fatty acids and proteins. Studies have shown that the bioactive compounds present in other parts of plant also, such as leaves, pods, roots stem etc. Maria Anwaar & Co- workers (2023) studied that Vigna mungo possesses pharmacological potential for the management of hyperthyroidism and improves estrus cycle disturbances. The study was designed to investigate the pharmacological potential of 70% methanolic extract of Vigna mungo for the management of hyperthyroidism and estrus cycle disturbances in female rats. The preliminary phytochemical analysis of the extract was performed. Hyperthyroidism was induced by administering levothyroxine (600 µg/kg; orally) for 14 days. Phytochemical analysis of the extract showed the presence of alkaloids, flavonoids, phenols, saponins, fats, and oils. The extract (300, 500, and 1,000 mg/kg; orally) and carbimazole (30 mg/kg; orally) were administered individually for the next 14 days. The extract showed dose-dependent antithyroid potential, as evidenced by the decrease in T<sub>3</sub> and T<sub>4</sub>, and the increase in TSH levels. Body weight, body temperature, fasting blood glucose levels, and prolonged diestrus phase were reversed towards normal and significant effects on the morphology of the thyroid gland and ovary of the rats were observed. Therefore, the findings reveal that V. mungo has the potential to treat experimentally-induced hyperthyroidism and estrus cycle disturbances.

## **Material And Method-**

This research aims to examine the impact of salinity stress on urad bean (*Vigna mungo*) seed germination by exposing seeds to different sodium chloride (NaCl) concentrations and analysing their germination responses. The study follows a structured experimental approach to ensure accuracy and reliability in data collection.

A total of five experiments were conducted which were identical in nature. Each of the five treatment groups consisted of two replicates, with 20 seeds per replicate, ensuring adequate sample size for statistical analysis. The inclusion of multiple salinity levels allows for a comprehensive evaluation of the effects of increasing salt concentrations on seed germination. Each treatment group (20 seeds of *Vigna mungo*) was then wrapped in damp paper towel and placed in five bowls separately. The five treatment groups were subjected to five different salinity concentration:

- 0mM/L NaCl (control) no salt stress, normal germination condition.
- 50mM/L NaCl (mild) low salt stress, minimal impact on germination and growth.
- 100mM/L NaCl (moderate) noticeable reduction in germination rate and seedling growth.

A MONTHLY, OPEN ACCESS, PEER REVIEWED (REFEREED) INTERNATIONAL JOURNAL Volume 04, Issue 08, August 2025

- 150mM/L NaCl (high)- significant inhibition on germination rate and seedling growth.
- 200mM/L NaCl (severe) severe reductions in germination, root and shoot growth, and psychological damage.

These concentrations were made using measured amounts of tap water and table salt, and are based upon the previous experiments to allow easier comparison between the results. Two tablespoons of these saline solutions were transferred to their respective bowls. The bowls were then closed leaving a little bit of space for aeration. Fresh supply of saline solutions was provided every other day. Number of germinated seeds were counted every day.

The experiment was designed using a completely randomized design (CRD), a widely accepted statistical method that ensures unbiased results. Seeds were divided into five treatment groups based on different salinity levels:

- 1. Control (0 mM NaCl): Seeds germinated under normal conditions without salt stress.
- 2. Low Salinity (50 mM NaCl): Seeds exposed to mild salinity stress.
- 3. Moderate Salinity (100 mM NaCl): Seeds subjected to moderate salinity stress.
- 4. High Salinity (150 mM NaCl): Seeds grown under high salinity stress.
- 5. Severe Salinity (200 mM NaCl): Seeds experiencing extreme salinity stress.

The bowls containing seeds were placed in a growth chamber under controlled environmental conditions to maintain uniformity across all treatments. The following conditions were maintained:

- **Temperature**:  $25 \pm 2$ °C (optimal temperature for urad bean germination)
- **Photoperiod**: 12-hour light / 12-hour dark cycle (to mimic natural germination conditions)
- **Relative Humidity**: 60–70% (to prevent excessive drying or fungal growth)

To maintain consistent salinity levels throughout the experiment, the paper towels were regularly moistened with the respective NaCl solutions.

The process of seed germination was monitored daily for 7 days, and several key germination parameters were recorded to assess the impact of salinity stress.

- Germination Percentage (GP): Defined as the proportion of seeds that successfully germinated under each treatment. This parameter provides a direct measure of the effect of salinity on the ability of urad bean seeds to initiate growth.
- Mean Germination Time (MGT): The average time required for germination, calculated using standard germination formulas. A longer MGT under saline conditions would indicate a delay in germination.
- Germination Rate Index (GRI): A measure of how quickly seeds germinate over time, calculated by considering the number of seeds germinating each day.
- 4.2 Seedling Growth Measurements
- The lengths of the radicle (root) and plumule (shoot) were measured on the 7th day using a digital calliper.
- The seedling vigour index (SVI) was determined using the formula:

#### [SVI=Germination Percentage×(Root length + Shoot length)]

This parameter helps evaluate the overall growth potential of the seedlings under different salinity treatments.

To determine whether salinity stress had a statistically significant effect on germination, the collected data was subjected to statistical analysis using ANOVA (Analysis of Variance) test. This method helps compare means across different treatment groups and identify variations in germination performance.

A MONTHLY, OPEN ACCESS, PEER REVIEWED (REFEREED) INTERNATIONAL JOURNAL Volume 04, Issue 08, August 2025

Additionally, Tukey's post-hoc test was performed to determine which specific salinity levels caused significant differences in germination. Data were expressed as mean  $\pm$  standard deviation (SD), and significance was determined at p < 0.05, meaning that observed differences were considered statistically meaningful if the probability of error was less than 5%.

We used www.icalcu.com, a free online data analysis calculator, to complete these two tests and generate relevant results.

#### **OBSERVATIONS**

### 1.Germination Percentage:

S. No.	NaCl conc. (mM/L)	Germination %
1.	Control	96.97
2.	50	91.54
3.	100	84.78
4.	150	73.65
5.	200	41.74

Germination percentage significantly declined with increasing NaCl concentrations. The control group (0 mM NaCl) exhibited the highest germination percentage, while the highest salinity level (200 mM NaCl) resulted in a drastic reduction in germination.

#### 2. Germination Rate and Mean Germination Time:

S. No.	NaCl conc. (mM/L)	Germination Rate (%)
1.	Control	80
2.	50	74
3.	100	63
4.	150	57
5.	200	49

These findings demonstrate that higher salinity levels not only decrease the total number of germinated seeds but also delay the germination process.

#### 3. Root and Shoot Length

Root and shoot length measurements indicated that salinity stress negatively affected seedling growth. The inhibition of root elongation was more severe than shoot growth, suggesting that salt stress primarily affects root development due to direct soil contact.

S. No.	NaCl Concentration (mM)	Root Length (cm)	Shoot Length (cm)
1.	Control	15.7	14.0
2.	50	12.8	11.2
3.	100	10.1	9.3
4.	150	8.9	6.8
5.	200	5.4	5.7

**Conclusion-** The present study aimed to assess the effect of salinity stress on the seed germination and early seedling growth of urad bean (*Vigna mungo*) an important pulse crop. Salinity is a significant abiotic stress

A MONTHLY, OPEN ACCESS, PEER REVIEWED (REFEREED) INTERNATIONAL JOURNAL Volume 04, Issue 08, August 2025

that affects plant growth by altering water availability, including ion toxicity, and causing oxidative stress. Based on the experimental findings, increasing salinity levels led to a decline in germination percentage, root and shoot length, and biomass accumulation. As a result, the energy required for metabolic processes during germination and early growth is reduced, leading to poor seedling growth. Higher NaCl (table salt) concentrations caused significant reduction in seedling growth, indicating that mung bean is sensitive to salinity stress, particularly above the level of 100mM/L.

#### References-

- 1. Anwaar M, Jabeen Q, Ali H, Nadeem A, Rasheed HMF (2023). *Vigna mungo* possesses pharmacological potential for the management of hyperthyroidism and improves estrus cycle disturbances. Bangladesh Journal of Pharmacology, Vol 18 No 3 (2023)
- 2. Arora, S., and Sharma, V. (2017). Reclamation and management of salt-affected soils for safeguarding agricultural productivity. J. Safe Agri. 1, 1–10.
- 3. Bradbury, M. and Ahmad, R. (1990). The effect of silicon on the growth of Prosopis juliflora growing in saline soil. Plant and Soil, 125: 71-74
- 4. Dhumal JS, Chaudhary SR, Chavan MJ (2019). A Review Bioactive Components of *Vigna mungo*. Journal of Drug Delivery and Therapeutics. 09 (4-s): 748-754
- 5. Dubey R.S. Rani M. (1990) Influence of NaCl salinization on the behaviour of protease, aminopeptidase and carboxypeptidase in rice seedlings in relation to salt tolerance. Australian Journal Plant Physiology, 17: 215-221
- 6. Flowers, T.J. and Flowers, S.A. (2005). Why does salinity pose such a difficult problem for plant breeders? Agric. Water Manag.
- 7.Food and Agriculture Organization of the United Nations (2015). Status World's Soil Resources; FAO: Rome, Italy, 650p, ISBN 978-92-5-109004-6 11. Gong, Y., Rao, L., and Yu, D. (2013). "Abiotic stress in plants," in Agricultural Chemistry, ed. M. Stoytcheva (Rijeka: InTech).
- 8. Jamil, M.; Lee, D.B.; Jung, K.Y.; Ashraf, M.; Lee, S.C.; Rha, E.S. Effect of salt (NaCl) stress on germination and early seedling growth of four Vegetable species. J. Cent. Eur. Agric. 2006, 7, 273–282.
- 9. Khan F, Nayab M, Ansari AN, Zubair M (2021) Medicinal Properties of Mash (*Vigna mungo* (Linn.) Hepper): A Comprehensive Review. Journal of Drug Delivery and Therapeutics. 2021; 11(3-S): 121-124
- 10. Kumar D. Kumar P. Deepika A. (2008) Effect of salinity on germination and early seedling growth of berseem. Environment and Ecology, 26(4C): 2369-2371.
- 11. Kumar S.A. Muthukumarasamy M. Panneerselvam R. (1996) Nitrogen metabolism in Black gram under NaCl stress. The Journal of the Indian Botanical Society, 75: 69-71.
- 12. Munns R. and Tester M. (2008). Mechanisms of salinity tolerance. Annu. Rev. Plant Biol. 59: 651–681.
- 13. Murillo-Amador B. Lopez-Aguilar R. Kaya C. Larrinaga-Mayoral J. Flores-Hernandez A. (2002) Comparative effects of NaCl and polyethylene glycol on germination, emergence and seedling growth of cowpea. Journal of Agronomy and Crop Science, 188: 235-247.
- 14. Paliwal, K. V. and Maliwal, G. L. (1980). Growth and nutrient uptake relationship of some crops in saline substrate. Ann. Arid Zone. 19: 251–25.
- 15. Sehrawat N., Jaiwal P.K., Yadav M., Bhat K.V. and Sairam R.K. (2013). Salinity stress restraining mung bean (*Vigna radiata* L. Wilczek) production: gateway for genetic improvement. Int. J. Agric. Crop Sci. 6: 505–509.

A MONTHLY, OPEN ACCESS, PEER REVIEWED (REFEREED) INTERNATIONAL JOURNAL Volume 04, Issue 08, August 2025

16. Tanji, K. K. (1990). "Nature and Extent of Agricultural Salinity," in Agricultural Salinity Assessment and Management, ASCE Manuals and Reports on Engineering Practice. No 71. ed K. K. Tanji (New York, NY: ASCE), 1–17. 27. Waisal, Y. (1972). Biology of Halophytes. Aca. Press, New York.